McMasters Technique & LA Parasite Ova

What you should accomplish during Lab.

1. After an introduction, students, working in pairs, will prepare and examine a McMasters Technique, Determine FEC and Determine % Efficacy

2. Be able to identify parasite ova & oocysts commonly found in ruminant, equine, and swine feces.

McMaster's Quantitation Technique

- 1) Fill McMaster's Graduated Vial (clear vial with 2 lines) to the bottom line with flotation solution (= 26 mls).
- 2) Add feces, about 4 gm, until the fluid level rises to the top line.
- 3) Pour this mixture into a clean beaker and mix thoroughly.
- 4) Pour mixture through a strainer into a 2nd clean beaker.
- 5) Mix strained mixture by pouring mixture from beaker to beaker a few times.
- 6) Withdraw a small amount of the well-mixed suspension with a pipette and load this into one side of the McMaster's counting chamber.
- 7) Mix suspension again by pouring mixture from beaker to beaker a few times.
- 8) Again, withdraw a small amount of the well-mixed suspension with a pipette and load the second side of the McMaster's counting chamber.
- 9) Wait 1 minute for eggs to rise to the top of the chamber.
- 10) Focus on the lines of the McMaster's chamber with 4X, then examine the chamber with 10X. (Scan for ova).
- 11) Examine the entire ruled area, counting all the eggs within the ruled areas
- 12) Add the total egg from each side of the chamber.
- 13) Multiply the sum of the 2 chambers by 25 to determine the eggs per gram (epg).
- Note: The McMaster's Quantitation Techniques is mainly for the quantitation
 of <u>Strongyle-type ova</u>, thus only strongyle-type eggs should be counted.
 However, a general idea (i.e. none, few, many...) of the number of other
 nematode ova, cestode ova & coccidian oocysts should be noted.
- Note: The McMaster's Chambers can NOT be examined with the 40X or 100X objectives.

Exercises

- 1. Fecal Egg Count & Fecal Floatation.
 - a) Perform a McMasters on the provided feces.
 - i. Count the number of strongyle-type eggs in each grid.
 - b) Determine and record the resulting FEC.

(Grid A: _____ + Grid B: _____) X 25 = ____ epg

c) Perform a Fecal Floatation with the strained fecal suspension.

- 2. % Efficacy
 - a) A fecal was collected at the time of deworming with Fenbendazole. At that time, a McMasters was performed and the pre-FEC was_____ epg. (pre-FEC will be provided by the instructor)
 - b) <u>Twelve days later</u> a Post-Treatment fecal was collected.

 Assume your results from today's lab is the Post-Treatment fecal.
 - c) Utilizing the % Efficacy formula and determine if the dewormer was effective or is showing resistance.

The formula for determining efficacy of an anthelmintic is:

i. % Efficacy of >95% is considered Efficacious

ii. % Efficacy of <95% is considered Not Efficacious.

Results

1.	Which fecal sample did you examin	ne:
		(Goat, Sheep, or Horse)
2.	The FEC: epg.	
3.	% Efficacy:% Resistance: Yes or No	
4 .	List the Parasite ova / oocysts th	nat you found.
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To Study

1. Observe specimens of parasite diagnostic stages presented on the overhead monitors.

• Utilize this lab and the <u>Parasitology Website</u> to learn these diagnostic stages as one will be responsible for identifying these on the Final Exam.

Ruminant

Oocysts: Eimeria.

Ova: Strongyle-type, Nematodirus, Strongyloides, Trichuris, Moniezia. Fasciola.

Equine

Ova: Strongyle-type, Strongyloides, Parascaris, Oxyuris, Anoplocephala.

Swine

Oocysts: Eimeria, Cystoisopora

Ova: Strongyle-type, Strongyloides, Ascaris, Trichuris

The Links for the Parasitology Website Diagnostic Keys are:

https://parasitology.cvm.ncsu.edu/vet900/m_keys.html https://parasitology.cvm.ncsu.edu/vet900/w_keys.html https://parasitology.cvm.ncsu.edu/vet900/quiz.html

Even though there are other parasites on the keys: <u>YOU ARE ONLY</u> RESPONSIBLE FOR THE DIAGNOSTIC STAGES LISTED ABOVE.

On The Exam you will be given the source of the specimen.

Example: "Dog Feces"